

Sertoli Cell Secreted Regulatory Factors

MICHAEL K. SKINNER

*Center for Reproductive Biology and Center for Integrated Biotechnology, School of Molecular Biosciences,
Washington State University, Pullman, Washington*

- I. INTRODUCTION
 - II. GROWTH FACTORS
 - III. HORMONES
 - IV. OTHER REGULATORY FACTORS
 - V. SUMMARY
- References

I. INTRODUCTION

The Sertoli cell provides the microenvironment and cytoarchitectural support for the developing spermatogenic cells and directly regulates the reproductive endocrinology of the male. This is in large part accomplished through the secretion of a wide variety of proteins. These secretory products include transport proteins to provide nutrient support to the germ cells, extracellular matrix and cell adhesion molecules to promote appropriate cell-cell interactions, and proteases to allow tissue remodeling during spermatogenesis. These Sertoli cell secreted factors can be categorized as nutritional factors (e.g., transport proteins) that support the nutrient requirements of the germ cells, environmental factors (e.g., extracellular matrix) that influence the physical content and extracellular environment between cells, and regulatory factors (e.g., growth factors) [1].

Regulatory factors are defined as factors that, through receptor-mediated signal transduction events, influence cellular function, growth, or differentiation on a molecular level. These regulatory factors can act

as autocrine agents to influence Sertoli cells or paracrine factors to influence neighboring cells such as spermatogenic cells, Leydig cells, or peritubular myoid cells [1]. These regulatory factors are essential for the careful control of testis development, spermatogenesis and male fertility, and male reproductive endocrinology. Two major subcategories of regulatory factors are growth factors and hormones.

Growth factors are defined as factors that influence cell proliferation and tissue growth. Although growth factors directly influence the cell cycle, growth factors can also have effects on a variety of other cellular functions and cell differentiation. Properties of a variety of growth factors are presented in Table 8.1. All the growth factors listed have been shown to be Sertoli cell secreted factors and are discussed in more detail later. These Sertoli cell secreted regulatory factors are all required for normal testis function and male fertility.

Another subcategory of Sertoli cell secreted regulatory factors is hormones. The local actions of hormone-like factors are often required to directly influence cellular function and differentiation. In contrast to growth factors, hormones generally do not directly influence cell proliferation. However, indirect effects mediated through the production of a growth factor do occur. These hormones are also needed to act as an endocrine agent at distant tissue and organs to regulate the reproductive endocrinology of the male. The positive and negative feedback systems built into endocrine regulation require the local production of hormones. Sertoli cell secreted hormones are discussed later.

TABLE 8.1 Properties of Several Common Growth Factors

| Growth factor | Abbreviation | Approximate size (kDa) | Examples of physiological action | Receptor(s) |
|--|--------------|------------------------|----------------------------------|--|
| Insulin-like growth factor I | IGF-I | 7.5 | Skeletal growth | IGF-I receptor |
| Insulin-like growth factor II | IGF-II | 7.5 | Fetal development | IGF-I and IGF-II |
| Epidermal growth factor | EGF | 6 | Tissue growth | EGF receptor |
| Transforming growth factor α | TGF α | 5 | Tissue growth | EGF receptor |
| Transforming growth factor β | TGF β | 25/dimer | Growth inhibition/tissue repair | TGF β , type 1, 2, and 3 receptors |
| Fibroblast growth factor | FGF | 17 | Angiogenesis/tissue growth | FGF receptor |
| Neurotrophins | NT-3 | 13 | Neuronal development | TrkC receptor |
| Interleukin 1 | IL-1 | 17 | Immune response/inflammation | IL-1 receptor |
| Kit ligand/stem cell factor | KL/SCF | 30 | Tissue growth/germ cells | KL receptor |
| Glial cell-derived neurotrophic factor | GDNF | 34 | Cell growth | Ret/GFR α receptor |

The initial review of this topic was published more than 10 years ago in *The Sertoli Cell* [2]. The advances since that time have been significant and will be the focus of the current review. Both Sertoli cell secreted growth factors and hormones are discussed. These secreted factors play a critical role in the development, growth, and maintenance of testis function. Sertoli cells are the principal cell regulating the process of spermatogenesis and these secreted regulatory factors are essential for Sertoli cell function.

II. GROWTH FACTORS

The development of the testis and maintenance of spermatogenesis require a precise growth regulation [3, 4]. All cell populations proliferate during embryonic testis development and in the early postnatal period. Sertoli cells become postmitotic and terminally differentiate during the early pubertal period once spermatogenesis is initiated [3–5]. The other somatic cells in the testis (e.g., Leydig, peritubular) have a slowed but continuous rate of growth in the adult [4–8]. Spermatogenic cells require a rapid rate of mitosis and meiosis at the onset of puberty that continues throughout adult life. The waves of spermatogenic cell proliferation require local control by Sertoli cells. The secretion of growth factors by Sertoli cells will have a role in the precise cell growth regulation required in the developing and adult testis.

A. Insulin-Like Growth Factors

Growth factors with structural similarity to insulin are in a family of factors termed insulin-like growth

factors (IGFs). These include IGF-I and IGF-II. IGF-I is a critical factor required for cell cycle progression and DNA synthesis in all proliferating cells. For this reason IGF-I is expressed and acts on nearly all cells. The serum level of IGF-I is high (e.g., 100 ng/mL) and is a principal liver product [9]. A family of IGF binding proteins exists to regulate the level of IGF bioactivity available to a cell and influence the homeostasis of the extracellular growth factors.

All testicular somatic cells express and respond to IGF-I including Sertoli cells [10–13]. IGF-I influences DNA synthesis and increases lactate and transferrin production by Sertoli cells [14, 15]. IGF-I also can influence the actions of FSH on Sertoli cells [16, 17] through unique signaling events [17]. Although the IGF-I produced by Sertoli cells [10–13] can act as an autocrine factor on the Sertoli cells, the high concentration of IGF-I in the interstitial fluid is available to the basal surface of the Sertoli cell. The blood–testis barrier created by Sertoli cells sequesters the developing spermatogenic cells. IGF-I available in the interstitium does not pass this barrier. Therefore, IGF-I secreted by Sertoli cells will likely have a critical role as a paracrine factor on germ cells (Fig. 8.1). Germ cells appear to be a site for IGF-I actions [18, 19]. Local production of IGF binding proteins by Sertoli cells has been shown to influence IGF-I actions and will contribute to this proposed Sertoli cell–germ cell interaction [12, 20, 21]. The ability of IGF-I produced by Sertoli cells to influence Leydig cell function has also been proposed [22]. Although these interactions can be observed *in vitro*, the physiological importance *in vivo* is questioned due to the levels of IGF-I and binding proteins in the interstitium [23]. It is likely Sertoli cell secreted IGF-I will be critical for spermatogenic cells, but actions on other

testis somatic cells is questioned due to other sources of IGF-I for these cells.

IGF-II has also been shown to mediate Sertoli cell–spermatogenic cell interactions [24–27]. The IGF-II receptor is present on both Sertoli cells and germ cells [25]. This receptor can also influence gene expression in the spermatogenic cells [26]. The IGF-II receptor is also termed the mannose-6-phosphate receptor and can bind a variety of additional ligands with appropriate carbohydrate specificity. Therefore, this IGF-II receptor system appears to mediate interactions between Sertoli cells and germ cells through a variety of potential ligands [27].

B. Transforming Growth Factor α

Transforming growth factor α (TGF α) is a member of the epidermal growth factor (EGF) family and binds to the same EGF receptor as EGF [28, 29]. TGF α is expressed as a transmembrane precursor that is proteolytically processed to release the mature peptide (Table 8.1). TGF α is secreted by a large number of normal cell types and promotes cell proliferation in normal and transformed cells.

EGF has been shown to influence spermatogenesis [30], and removal of serum levels of EGF reduces sperm numbers [31]. Circulating concentrations of EGF, however, are thought to be too low to have physiological influences [32]. EGF is not expressed in the rodent testis or in Sertoli cells [33]. In contrast, TGF α is expressed by Sertoli cells [33]. Both peritubular myoid cells and Leydig cells also produce TGF α [33, 34]. Peritubular cells and

Leydig cells proliferate in response to TGF α [33]. TGF α has been shown to influence Sertoli cell lactate and estrogen production [35] and can influence Sertoli cell DNA synthesis if the cells are derived from prepubertal animals [36, 37]. TGF α also influences transferrin production by Sertoli cells [38]. The actions of TGF α on Sertoli cells and the testis are dependent on the developmental stage. Embryonic testis growth requires TGF α expression and action on most cell types [4]. At the onset of puberty, the Sertoli cells differentiate and become postmitotic. Sertoli cell responsiveness to TGF α /EGF declines as the adult stage of development is obtained [39]. Other cell types continue to utilize TGF α for growth regulation. The spermatogenic cells also utilize TGF α , and Sertoli cell–derived TGF α is postulated to be an important paracrine interaction of this Sertoli cell regulatory factor [40] (Fig. 8.1). Sertoli production of TGF α is also capable of influencing adjacent peritubular cells [39, 40].

C. Transforming Growth Factor β

Transforming growth factor β (TGF β) has three mammalian isoforms (TGF β_1 , TGF β_2 , and TGF β_3) that act as inhibitors of cell proliferation and facilitate cellular differentiation in a wide variety of cells [41]. TGF β acts by inhibiting the actions of growth stimulators (e.g., TGF α) and promoting the expression of extracellular matrix components. Most cell types contain the receptors for TGF β isoforms. Although found in transformed cells, TGF β regulates most normal cell types [41].

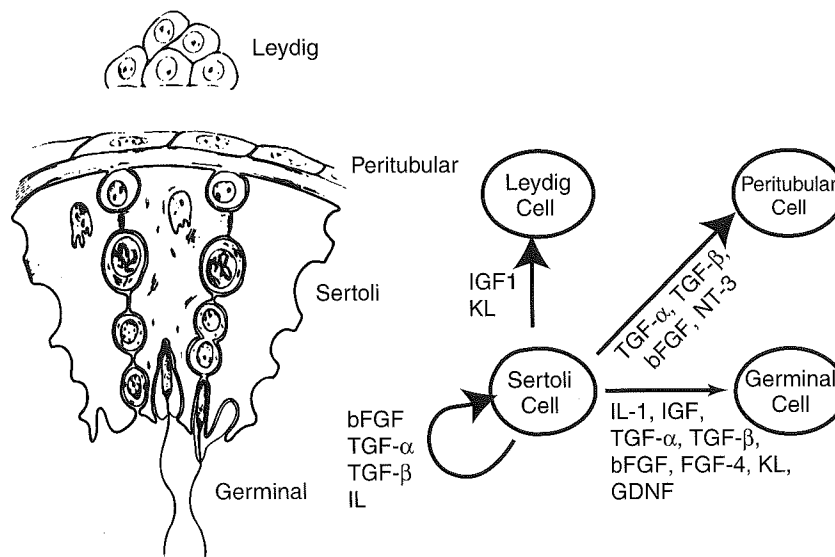


FIGURE 8.1. Sertoli cell secreted regulatory factors.

TGF β isoforms are expressed by Sertoli cells [40, 42, 43] and Sertoli cells are one of the few tissues that express TGF β 3 [40, 44]. Sertoli cells express all three isoforms under distinct hormonal regulation [40, 45]. The developmental regulation of TGF β expression in the testis of the different isoforms suggests distinct functions and/or differential regulation [45, 46]. TGF β expression by Sertoli cells is critical for embryonic testis development [46, 47]. At the onset of puberty, TGF β expression by Sertoli cells also corresponds to alterations in cell growth [45]. TGF β ₁ is high prepubertally and then declines to low levels in the adult. TGF β ₂ expression is similar and FSH appears to promote the decrease in expression by Sertoli cells [45]. TGF β ₃ has a transient increase in expression at the onset of puberty [45].

The site of action for Sertoli cell secreted TGF β is suggested from the analysis of TGF β receptor localization. All cell types in the testis contain type I and II TGF β receptors [48–50], including spermatogenic cells. Peritubular cells produce and respond to TGF β with a decline in cellular proliferation induced by TGF α [42, 45]. Peritubular cells also produce more extracellular matrix components in response to TGF β [42]. TGF β also induce peritubular cell contractility that is needed for sperm transport in the seminiferous tubule [51]. TGF β does not have major effects on Sertoli cell proliferation or functional genes, but does increase production of plasminogen activator [52]. TGF β also appears to influence the tight junctions between Sertoli cells [53, 54]. Leydig cells are also a site of TGF β production and action. TGF β inhibits Leydig cell steroidogenesis [55–57]. Therefore, critical targets for the Sertoli cell secreted TGF β isoforms are spermatogenic cells and peritubular cells (Fig. 8.1). Abnormalities in TGF β expression have been suggested as causal in testicular diseases such as fibrosis [58]. The bioactivity of TGF β produced by Sertoli cells is also a factor in the function of TGF β [59]. TGF β also can be inhibited by specific proteins such as Bambi [60], which can influence the actions of the Sertoli cell expressed TGF β . TGF β is a critical regulatory factor produced by Sertoli cells to influence testis development, function, and spermatogenesis.

D. Fibroblast Growth Factor

Several members of the fibroblast growth factor (FGF) family have been shown to influence testis function. FGF family members regulate cellular growth and differentiation, as well as tissue angiogenesis [61]. FGFs have widespread tissue distribution and each of the family members has distinct receptors and specific functions.

Basic FGF (bFGF) is one of the most studied FGF family members. Initially bFGF was identified in

whole testis [62, 63] and subsequently was found to be expressed by Sertoli cells [64, 65]. bFGF production by Sertoli cells was found to be increased by FSH [65]. bFGF has been shown to stimulate Sertoli cell growth prepubertally [66]. The ability of FSH to stimulate Sertoli cell growth prepubertally may in part be indirectly mediated through bFGF production [65]. Early postnatal actions of FGF also suggest potential indirect effects on gonocyte proliferation [67]. Sertoli cells also respond to bFGF by an increase in the production of proteoglycans [68], lactate, and glucose metabolism [69]. Localization of the bFGF receptor (FGFR type 1) demonstrated receptors in Sertoli cells, Leydig cells, peritubular cells, and germ cells [70]. Therefore, the actions of bFGF secretion by Sertoli cells could mediate cellular interactions with spermatogenic cells and peritubular cells, as well as autocrine actions on Sertoli cells (Fig. 8.1).

Another member of the FGF family, Hst-1/FGF-4, has been shown to be expressed by Sertoli cells [71]. A conditional knockout of FGF-4 resulted in impaired fertility and a knockin caused enhanced spermatogenesis [72]. This observation suggests a potential Sertoli cell–spermatogenic cell interaction mediated by FGF-4 (Fig. 8.1). The overexpression of FGF-4 was also found to protect the testis from the chemotherapeutic drug adriamycin [72]. Therefore, Sertoli cell expression of FGF-4 appears to be important for normal testis function.

FGF-9 is another member of the FGF family that has been shown to be important for early embryonic testis development [73]. FGF-9 appears to be a downstream gene to Sry and is expressed by Sertoli cells to influence male sex differentiation through the adjacent mesenchymal tissue. A knockout of FGF-9 results in sex reversal and impaired testis development [73]. Functions of FGF-9 later in development have not been thoroughly investigated.

E. Neurotrophins

The neurotrophin family of growth factors includes nerve growth factor (NGF), neurotrophin 3 (NT-3), neurotrophin 4/5 (NT-4/5), and brain-derived neurotrophic factor (BDNF). The high-affinity receptors for these factors are the trk receptors and the low-affinity ones are the LNGFR. Although these neurotrophins are critical for neurons and associated cells, they also have been shown to be expressed by a number of non-neuronal tissues including the testis.

NGF has been the most highly investigated neurotrophin in the testis. NGF appears in the adult testis to primarily be expressed by spermatogenic cells [74–76], whereas the NGF receptor trkA is expressed in Sertoli cells [76–78]. The NGF precursor protein is highly expressed in round spermatids, while the NGF acts to

promote Sertoli cell viability [76] and cellular functions such as lactate and estrogen production [78]. Although some studies have suggested NGF expression in the adult Sertoli cell [79], the majority have suggested NGF mediates a spermatogenic cell–Sertoli cell interaction and adult Sertoli cells do not express NGF [76–78, 80]. In contrast to adult Sertoli cells, NGF does appear to be expressed by late embryonic and early postnatal Sertoli cells [81]. The NGF receptor (trkA) was expressed in Sertoli cells, peritubular cells, and interstitial cells at these stages of development [81]. Therefore, NGF is a Sertoli cell secreted growth factor, but its expression by Sertoli cells appears to be more important in the perinatal period than in the adult.

Other neurotrophins expressed in the adult testis have not been extensively studied. NT-3 was found to localize in spermatogenic cells, whereas BDNF may be expressed by adult Sertoli cells [82]. In contrast, BDNF is not expressed in the embryonic or postnatal testis [81]. More thorough investigation of other neurotrophins is needed to clarify roles in the adult testis. Observations suggest that Sertoli cell expressed neurotrophins play a critical role during development, but in the adult, spermatogenic cells influence Sertoli cells through neurotrophin production.

NT-3 appears to be a critical neurotrophin secreted by Sertoli cells during the initial stages of male sex determination and testis development [81, 83]. At the time of Sertoli cell fate differentiation by the testis determining factor Sry, Sertoli cells express NT-3 as a potential immediate downstream gene to Sry [81, 83]. This Sertoli cell secreted NT-3 acts on adjacent mesonephros cells as a chemotactic factor and promotes migration into the testis, and the migrating mesonephros cells become peritubular cells and promote seminiferous cord formation, which is the first morphological event in testis development [81, 83, 84]. The NT-3 receptor trkC is on the migrating mesonephros cells and blocking the actions of this receptor inhibits cord formation and testis development [83]. This has also been recently confirmed in the human fetal testis [85]. NT-3 also can act as a direct chemotactic agent to induce mesonephros migration into the developing testis [84]. Observations suggest Sertoli cell secretion of NT-3 is a downstream event to Sry and critical to early embryonic testis development by acting on early developing peritubular cells. In contrast, adult Sertoli cells do not appear to express NT-3, such that this is primarily an embryonic function of Sertoli cells.

F. Cytokines

Cytokines are defined as growth factors involved in immune cell communication and the immune system.

However, these proteins also influence nonimmune cells and tissues. A number of different cytokines that have been shown to have functions in the testis are outlined next.

The interleukins (ILs) are a family of cytokines produced by activated lymphocytes and macrophages. Interleukin 1 beta (IL-1 β) is secreted by lymphocytes, whereas interleukin 1 alpha (IL-1 α) is produced by a wide variety of nonimmune tissues. IL-1 α has been shown to be expressed by Sertoli cells [86] and appears to influence spermatogenic cells [87]. The expression of IL-1 α by Sertoli cells appears dependent on the presence of germ cells [88]. IL-1 α can directly influence prepubertal Sertoli cell growth [89], transferrin gene expression [90], and lactate production [91]. Interestingly, Sertoli cells also express an antagonist form of IL-1 α termed IL-1ra [92, 93]. These positive and negative acting forms of IL-1 α are proposed to have a role in regulating cellular interactions between Sertoli cells and germ cells [92–94]. In contrast to IL-1 α , IL-1 β is not expressed by Sertoli cells [92, 93]. However, IL-1 β can induce the expression of IL-1 α by Sertoli cells [95].

In addition to IL-1 α , Sertoli cells produce interleukin 6 (IL-6) in response to the autocrine actions of IL-1 α [96–98]. Residual bodies induce Sertoli cell expression of IL-1 α that in an autocrine manner induces Sertoli cell production of IL-6. Analysis of the hormonal regulation of IL-1 and IL-6 demonstrates distinct regulation [99] and secretion into different tubule compartments [100]. Although autocrine actions of IL-6 on Sertoli cells have been postulated [101, 102], an important function of Sertoli cell secreted interleukins is communication with the immune cells in the interstitial compartment [103, 104]. Leydig cells also produce these interleukins, probably for a similar function [105, 106].

Other cytokines produced by Sertoli cells include interferon α (IFN- α) [107] and α_2 -macroglobulin (α_2 -MG) [108]. Both are proposed to interact with immune cells in the interstitium and possibly mediate other cell–cell interactions. Macrophage populations in the interstitium also are targets for macrophage migration inhibiting factor (MIF), which is primarily produced by Leydig cells [109, 110], but under appropriate conditions is also produced by Sertoli cells [109]. These and other cytokines [111] are likely to promote critical interactions with immune cells and Sertoli cell secreted cytokines will have a role in these cellular interactions.

G. Kit Ligand/Stem Cell Factor

Kit ligand (KL), also known as stem cell factor (SCF), acts at the kit receptor and is known to be important

for primordial germ cell viability and cell migration to the embryonic gonad. KL is required for normal germ cell proliferation and fertility of both the male and female. Sertoli cells are known to secrete KL and the spermatogonia express the kit receptor [112, 113]. Inhibition of KL reduces spermatogenesis and specifically the early mitotic events of germ cell development. Therefore, KL produced by Sertoli cells is needed in the embryo for germ cell survival and proliferation, early postnatally for gonocyte development [114], and in the adult for spermatogonia proliferation. This is a classic Sertoli cell–spermatogenic cell paracrine interaction in that the growth factor is produced only by one cell and receptor is in the target germ cell (Fig. 8.1). Gonadotropins regulate the expression of KL by Sertoli cells and various agents such as gonadotropin-releasing hormone (GnRH) [115], growth-hormone-releasing hormone (GHRH) [116], and somatostatin [117] can influence KL production. The physiological importance of this Sertoli cell secreted factor is shown in forms of male infertility. Impaired expression of kit [118] and mutated forms of KL [119] result in subfertility in human infertility patients. In addition to the Sertoli cell produced KL acting on spermatogonia, Leydig cells have been shown to potentially be a target during development or regeneration of the testis [120, 121].

H. Glial Cell–Derived Neurotrophic Factor

Glial cell–derived neurotrophic factor (GDNF) is a member of the TGF β superfamily and neurotrophin family. GDNF has been shown to be secreted by Sertoli cells and acts on spermatogonia through the ret/GFR receptor specifically located on spermatogonia [122]. GDNF appears to promote spermatogonia stem cells to initiate development [122, 123]. Spermatogonia cell proliferation is influenced by GDNF [124, 125]. A related growth factor, neurturin, also has similar functions [125]. Therefore, similar to KL, GDNF is secreted by Sertoli cells to act on early-stage spermatogonia to promote cell proliferation and development (Fig. 8.1). Disruption of GDNF expression alters spermatogenesis [126] and overexpression of GDNF causes a non-metastatic germ cell tumor by hyperproliferation of spermatogonia [127]. Although GDNF can influence Sertoli cell proliferation prepubertally, the spermatogonia cell is the primary target in the adult [128].

III. HORMONES

Sertoli cells are endocrine cells in that they respond to hormones (e.g., FSH and androgen) and also produce hormones. In addition to the secretion of growth

factors by Sertoli cells, hormones are also important regulatory factors secreted. The major hormones produced by Sertoli cells are reviewed next.

A. Inhibin/Activin

More than 70 years ago, Sertoli cells were shown to produce a regulatory agent termed inhibin [129] that inhibits FSH production by the pituitary [130, 131]. The two inhibin gene product subunits α and β form a dimer for inhibin, whereas a β homodimer produces activin that can stimulate FSH production. Both inhibin and activin are now known to have a wide variety of biological functions and are produced by a number of tissues. Sertoli cells under the control of FSH produce inhibin [132–135]. Although the major role of inhibin is to feed back information to the pituitary to regulate FSH production, local actions within the testis are also known [136, 137]. Sertoli cells can respond to activin to influence postnatal Sertoli cell proliferation [138]. Inhibin may influence germ cells [139, 140] and germ cells can influence the expression of inhibin by Sertoli cells [141]. The cellular interactions mediated by inhibin and activin in the testis are complicated by their expression by multiple cell types. Leydig cells can produce both inhibin and activin [142–145]. Recently, activin has been shown to be produced by peritubular cells and may regulate Sertoli cells through a paracrine interaction [146, 147]. Therefore, the specific paracrine and autocrine roles for inhibin and activin remain to be fully elucidated. However, the endocrine roles for these factors are better established. Abnormalities in inhibin levels correlating with abnormal pubertal development and testicular dysfunction have been documented [148–150].

B. Müllerian Inhibiting Substance

Müllerian inhibiting substance (MIS), also termed anti-Müllerian hormone (AMH), is secreted by embryonic Sertoli cells and promotes the regression of the Müllerian duct and female reproductive tract development [151, 152]. MIS was first identified in the fetal and neonatal testes [153] and was subsequently cloned and found to be expressed by Sertoli cells [154–156]. MIS is a TGF β family member. MIS expression is induced upon male sex determination by Sertoli cells and then declines to negligible levels in the adult [157, 158]. FSH inhibits the expression of MIS correlating with the decline during pubertal testis development [159]. MIS acts at specific receptors [151] to promote regression of the Müllerian duct and to regulate androgen production by Leydig cells [160]. The regulation of MIS expression has been examined on the transcriptional level

and demonstrated that the SF1, GATA-4, and Sox8 transcription factors regulate the MIS promoter [161–163]. FSH, thyroid hormone, and androgen can regulate the expression of MIS in Sertoli cells [164, 165]. MIS/AMH appears to be a Sertoli cell secretory factor that has a critical role during embryonic development as an endocrine hormone. Genetic mutations in MIS or its receptor cause human mutations in premature developmental defects, persistent Müllerian ducts, and azoospermia [166–168].

C. Estrogen

Sertoli cells express aromatase prepubertally so they can utilize androgen production by Leydig cells to produce estrogen. As Sertoli cells differentiate, aromatase expression drops such that negligible estrogen is produced in the adult testis. The knockout null mutations in the estrogen receptor (ER) demonstrated alterations in spermatogenesis and infertility [169]. The phenotype was found to be primarily mediated through rat testis and efferent duct abnormalities [170, 171]. However, estrogen production by Sertoli cells appears to also have direct effects on testis function and spermatogenesis [172]. Localization of the ER α demonstrated expression in Leydig cells and spermatogenic cells [173], whereas ER β was found in Sertoli cells in the adult stage and in the fetal stage in Sertoli, Leydig, peritubular, and germ cells [174]. Estrogen can act on Sertoli cells and influence FSH actions [175]. As stated, FSH inhibits estrogen production and IGF-1 can alter these effects [16]. Sertoli cell secretion of estrogen appears to be primarily a role for the prepubertal period and as the testis develops in response to FSH, estrogen production becomes negligible. Estrogen may have a role in suppressing prepubertal androgen produced by Leydig cells and may influence germ cells at later stages. Specific roles for estrogen in the adult remain to be elucidated.

IV. OTHER REGULATORY FACTORS

A. Uncharacterized Factors

Several regulatory factors have been postulated to be produced by Sertoli cells and have specific roles in mediating interactions with other cell types. These factors have not been characterized beyond their biological activity. The first was seminiferous growth factor (SGF), which had mitogenic activity and was distinct from FGF and stimulated proliferation of several Sertoli cell lines [176–178]. The second was

Sertoli cell secreted growth factor (SCSGF), which had some similarities to TGF α and also stimulated the growth of several Sertoli cell lines [179]. These factors may well be some of the factors described earlier and have similar biological activities.

A number of studies have suggested that Sertoli cells secrete a factor(s) that influences Leydig cell function and steroidogenesis. Abnormal physiological and environmental insults that affect Sertoli cell function have been shown to indirectly influence Leydig cell function [180–192]. Sertoli cell conditioned medium has been used to identify factors that stimulate cell proliferation and suppress steroidogenesis [193]. The evidence for such an activity is very convincing, but the factor(s) have not been characterized. Again a number of the Sertoli cell secreted factors mentioned earlier have this activity on Leydig cells (e.g., TGF α , bFGF).

Sertoli cells have also been shown to produce factors such as erythropoietin [194], leukemia inhibitory factor (LIF), and ciliary neurotrophic factor (CNTF) [195], but the specific expression and actions of these factors needs to be assessed. Recently, the genomic microarray analysis of Sertoli cell gene expression has suggested the expression of a number of secreted regulatory factors [196]. Examination of these factors primarily provided the same list as that discussed earlier. It is likely that new factors secreted by Sertoli cells will be determined, but a more genomic approach should facilitate the identification and initial detection versus the more classic protein biochemistry approaches used in the past.

B. Lipids

In addition to protein regulatory agents, nonprotein factors such as lipids are also found to have dramatic biological activities. Although many lipids have been primarily localized within the cell, some lipids such as lysophospholipids and thromboxanes are soluble and are known to bind specific G-protein coupled receptors on the surface of cells [197, 198]. These lipid-induced receptor-mediated events alter cellular functions in a similar manner to protein growth factors [197]. The testis has unique forms of metabolic enzymes generating these lipids [199–201], such that lipid factors may be secreted by Sertoli cells. Sertoli cells have been shown to produce soluble sphingosine molecules and other lysophospholipids [202–204]. The specific lipids secreted and functions remain to be elucidated, but due to the important functions of these extracellular soluble lipids it is likely they will be important regulators of testis function and spermatogenesis.

V. SUMMARY

The secretion of regulatory factors by Sertoli cells is critical for normal testis development and function. Abnormalities in the expression or action of these factors often result in infertility or endocrine defects. As shown in Figure 8.1 and Table 8.2, a number of growth factors are secreted by Sertoli cells to influence spermatogenic cells. Because Sertoli cells are the principal regulatory cells for spermatogenesis, it is not surprising that many of the growth factor's primary targets are germ cells. Since the original review of this topic, several major observations have been made regarding Sertoli cell secreted growth factors [2]. Knowledge that the production of KL and GDNF by Sertoli cells directly regulates spermatogonia cell proliferation and development represents a significant advance in our understanding of the control of the initiation of the spermatogenic wave and regulation of the spermatogenic stem cell population. Recently, human male infertility conditions have been linked to abnormalities in the expression and action of these factors. Other specific factors that have the characteristics of the growth factor being expressed by Sertoli cells and actions only on a specific stage of germ cells are likely to be identified. The number of

TABLE 8.2 Sertoli Cell Secreted Growth Factors

| Growth factor | Site of action | Function |
|---|--|---|
| Insulin-like growth factor I (IGF-I) | Germ cells Sertoli cells Leydig cells | Cell cycle progression and growth (i.e., S-phase DNA synthesis) |
| Insulin-like growth factor II (IGF-II) | Germ cells | Cell-cell signaling |
| Transforming growth factor α (TGF α) | Germ cells Sertoli cells Peritubular cells | Cell growth initiation |
| Transforming growth factor β (TGF β) | Germ cells Sertoli cells Peritubular cells | Growth inhibition and increase in cell differentiated function |
| Basic fibroblast growth factor (bFGF) | Germ cells Sertoli cells Peritubular cells | Cell growth initiation |
| Neurotrophin 3 (NT-3) | Peritubular cells | Cell migration (i.e., mesonephros) and growth |
| Interleukin 1 and 6 (IL-1 and IL-6) | Germ cells | Cell-cell signaling |
| Kit ligand/stem cell factor (KL/SCF) | Germ cells Leydig cells | Cell growth (i.e., spermatogonial) |
| Glial cell-derived neurotrophic factor (GDNF) | Germ cells | Cell growth (i.e., spermatogonial stem cells) |

TABLE 8.3 Sertoli Cell Secreted Hormones

| Hormone | Site of action | Function |
|---|----------------|--|
| Inhibin | Pituitary | Reduce FSH production |
| Müllerian inhibiting substance/anti-Müllerian hormone (MIS/AMH) | Müllerian duct | Regress female reproductive tract development |
| Estrogen | Leydig cells | Reduce steroidogenesis and cell-cell signaling |

growth factors listed in Table 8.2 will likely grow further during the next decade as well. The microenvironment within the testis and local cell-cell interactions mediated by regulatory factors such as those listed in Table 8.2 require elucidation for us to understand how hormones regulate testis function and how therapeutic treatments for male infertility can be developed.

The Sertoli cells also secrete several critical hormones, as listed in Table 8.3. Sertoli cells are critical to male reproductive endocrinology because they are a target for several hormones (e.g., FSH and testosterone) and they produce hormones. Feedback pathways are always required in a biological process and endocrine system, so it is not surprising that a critical inhibitory feedback protein like inhibin is produced. In addition, the negative feedback estrogen provides prepubertally to suppress Leydig cell androgen production is also anticipated. The information that has developed during the past decade [2] indicates that these factors also have other roles in mediating regulatory interactions between cells within the testis. Although the specific functions of the local actions of the inhibin, activin, and estrogen need to be elucidated, clearly they have local paracrine functions independent of the endocrine role of these factors. This will likely be an intense area of research for a number of years.

References

- Skinner, M. K. (1991). Cell-cell interactions in the testis. *Endocr. Rev.* **12**(1), 45-77.
- Skinner, M. K. (1993). Secretion of growth factors and other regulatory factors. In "The Sertoli Cell" (M. D. Griswold and L. D. Russell, eds.), pp. 237-247. Cache River Press, Clearwater, FL.
- Clermont, Y., and Perey, B. (1957). Quantitative study of the cell population of the seminiferous tubules in immature rats. *Am. J. Anat.* **100**(2), 241-267.
- Levine, E., Cupp, A. S., Miyashiro, L., and Skinner, M. K. (2000). Role of transforming growth factor-alpha and the epidermal growth factor receptor in embryonic rat testis development. *Biol. Reprod.* **62**(3), 477-490.
- Orth, J. M. (1982). Proliferation of Sertoli cells in fetal and postnatal rats: a quantitative autoradiographic study. *Anat. Rec.* **203**(4), 485-492.

6. Teerds, K. J., De Rooij, D. G., Rommerts, F. F., van der Tweel, I., and Wensing, C. J. (1989). Turnover time of Leydig cells and other interstitial cells in testes of adult rats. *Arch. Androl.* **23**(2), 105–111.
7. Lording, D. W., and De Kretser, D. M. (1972). Comparative ultrastructural and histochemical studies of the interstitial cells of the rat testis during fetal and postnatal development. *J. Reprod. Fertil.* **29**(2), 261–269.
8. Hardy, M. P., Zirkin, B. R., and Ewing, L. L. (1989). Kinetic studies on the development of the adult population of Leydig cells in testes of the pubertal rat. *Endocrinology* **124**(2), 762–770.
9. Daughaday, W. H., and Rotwein, P. (1989). Insulin-like growth factors I and II. Peptide, messenger ribonucleic acid and gene structures, serum, and tissue concentrations. *Endocr. Rev.* **10**(1), 68–91.
10. Chatelain, P. G., Naville, D., and Saez, J. M. (1987). Somatomedin-C/insulin-like growth factor 1-like material secreted by porcine Sertoli cells *in vitro*: characterization and regulation. *Biochem. Biophys. Res. Commun.* **146**(3), 1009–1017.
11. Smith, E. P., Svoboda, M. E., Van Wyk, J. J., Kierszenbaum, A. L., and Tres, L. L. (1987). Partial characterization of a somatomedin-like peptide from the medium of cultured rat Sertoli cells. *Endocrinology* **120**(1), 186–193.
12. Cailleau, J., Vermeire, S., and Verhoeven, G. (1990). Independent control of the production of insulin-like growth factor I and its binding protein by cultured testicular cells. *Mol. Cell Endocrinol.* **69**(1), 79–89.
13. Naville, D., Chatelain, P. G., Avallet, O., and Saez, J. M. (1990). Control of production of insulin-like growth factor I by pig Leydig and Sertoli cells cultured alone or together. Cell–cell interactions. *Mol. Cell Endocrinol.* **70**(3), 217–224.
14. Oonk, R. B., Jansen, R., and Grootegoed, J. A. (1989). Differential effects of follicle-stimulating hormone, insulin, and insulin-like growth factor I on hexose uptake and lactate production by rat Sertoli cells. *J. Cell Physiol.* **139**(1), 210–218.
15. Skinner, M. K., and Griswold, M. D. (1983). Multiplication stimulating activity (MSA) can substitute for insulin to stimulate the secretion of testicular transferrin by cultured Sertoli cells. *Cell Biol. Int. Rep.* **7**(6), 441–446.
16. Rappaport, M. S., and Smith, E. P. (1996). Insulin-like growth factor I inhibits aromatization induced by follicle-stimulating hormone in rat Sertoli cell culture. *Biol. Reprod.* **54**(2), 446–452.
17. Khan, S. A., Ndjountche, L., Pratchard, L., Spicer, L. J., and Davis, J. S. (2002). Follicle-stimulating hormone amplifies insulin-like growth factor I-mediated activation of AKT/protein kinase B signaling in immature rat Sertoli cells. *Endocrinology* **143**(6), 2259–2267.
18. Handelsman, D. J., Spaliviero, J. A., Scott, C. D., and Baxter, R. C. (1985). Identification of insulin-like growth factor-I and its receptors in the rat testis. *Acta. Endocrinol. (Copenhagen)* **109**(4), 543–549.
19. Tres, L. L., Smith, E. P., Van Wyk, J. J., and Kierszenbaum, A. L. (1986). Immunoreactive sites and accumulation of somatomedin-C in rat Sertoli-spermatogenic cell co-cultures. *Exp. Cell Res.* **162**(1), 33–50.
20. Rappaport, M. S., and Smith, E. P. (1995). Insulin-like growth factor (IGF) binding protein 3 in the rat testis: follicle-stimulating hormone dependence of mRNA expression and inhibition of IGF-I action on cultured Sertoli cells. *Biol. Reprod.* **52**(2), 419–425.
21. Besset, V., Le Magueresse-Battistoni, B., Collette, J., and Benahmed, M. (1996). Tumor necrosis factor alpha stimulates insulin-like growth factor binding protein 3 expression in cultured porcine Sertoli cells. *Endocrinology* **137**(1), 296–303.
22. Lejeune, H., Sanchez, P., and Saez, J. M. (1998). Enhancement of long-term testosterone secretion and steroidogenic enzyme expression in human Leydig cells by co-culture with human Sertoli cell-enriched preparations. *Int. J. Androl.* **21**(3), 129–140.
23. Zhou, J., and Bondy, C. (1993). Anatomy of the insulin-like growth factor system in the human testis. *Fertil. Steril.* **60**(5), 897–904.
24. O'Brien, D. A., Gabel, C. A., Rockett, D. L., and Eddy, E. M. (1989). Receptor-mediated endocytosis and differential synthesis of mannose 6-phosphate receptors in isolated spermatogenic and Sertoli cells. *Endocrinology* **125**(6), 2973–2984.
25. O'Brien, D. A., Welch, J. E., Fulcher, K. D., and Eddy, E. M. (1994). Expression of mannose 6-phosphate receptor messenger ribonucleic acids in mouse spermatogenic and Sertoli cells. *Biol. Reprod.* **50**(2), 429–435.
26. Tsuruta, J. K., and O'Brien, D. A. (1995). Sertoli cell-spermatogenic cell interaction: the insulin-like growth factor-II/cation-independent mannose 6-phosphate receptor mediates changes in spermatogenic cell gene expression in mice. *Biol. Reprod.* **53**(6), 1454–1464.
27. Tsuruta, J. K., Eddy, E. M., and O'Brien, D. A. (2000). Insulin-like growth factor-II/cation-independent mannose 6-phosphate receptor mediates paracrine interactions during spermatogonial development. *Biol. Reprod.* **63**(4), 1006–1013.
28. Derynck, R. (1988). Transforming growth factor alpha. *Cell* **54**(5), 593–595.
29. Carpenter, G., and Cohen, S. (1990). Epidermal growth factor. *J. Biol. Chem.* **265**(14), 7709–7712.
30. Stastny, M., and Cohen, S. (1972). The stimulation of ornithine decarboxylase activity in testes of the neonatal mouse. *Biochem. Biophys. Acta.* **261**(1), 177–180.
31. Tsutsumi, O., Kurachi, H., and Oka, T. (1986). A physiological role of epidermal growth factor in male reproductive function. *Science* **233**(4767), 975–977.
32. Carpenter, G., and Zendegui, J. (1986). A biological assay for epidermal growth factor/urogastrone and related polypeptides. *Anal. Biochem.* **153**(2), 279–282.
33. Skinner, M. K., Takacs, K., and Coffey, R. J. (1989). Transforming growth factor-alpha gene expression and action in the seminiferous tubule: peritubular cell–Sertoli cell interactions. *Endocrinology* **124**(2), 845–854.
34. Teerds, K. J., Rommerts, F. F., and Dorrington, J. H. (1990). Immunohistochemical detection of transforming growth factor-alpha in Leydig cells during the development of the rat testis. *Mol. Cell Endocrinol.* **69**(1), R1–6.
35. Mallea, L. E., Machado, A. J., Navaroli, F., and Rommerts, F. F. (1986). Epidermal growth factor stimulates lactate production and inhibits aromatization in cultured Sertoli cells from immature rats. *Int. J. Androl.* **9**(3), 201–208.
36. Petersen, C., Froysa, B., Boitani, C., and Soder, O. (2000). Transforming growth factor-alpha stimulates Sertoli cell proliferation *in vitro*. *Andrologia* **32**(1), 62–63.
37. Petersen, C., Boitani, C., Froysa, B., and Soder, O. (2001). Transforming growth factor-alpha stimulates proliferation of rat Sertoli cells. *Mol. Cell Endocrinol.* **181**(1–2), 221–227.
38. Onoda, M., and Suarez-Quian, C. A. (1994). Modulation of transferrin secretion by epidermal growth factor in immature rat Sertoli cells *in vitro*. *J. Reprod. Fertil.* **100**(2), 541–550.
39. Mullaney, B. P., and Skinner, M. K. (1992). Transforming growth factor-alpha and epidermal growth factor receptor gene expression and action during pubertal development of the seminiferous tubule. *Mol. Endocrinol.* **6**(12), 2103–2113.
40. Mullaney, B. P., and Skinner, M. K. (1991). Growth factors as mediators of testicular cell–cell interactions. *Baillieres Clin. Endocrinol. Metab.* **5**(4), 771–790.
41. Roberts, A. B., and Sporn, M. B. (1988). Transforming growth factor beta. *Adv. Cancer Res.* **51**, 107–145.
42. Skinner, M. K., and Moses, H. L. (1989). Transforming growth factor beta gene expression and action in the seminiferous

- tubule: peritubular cell-Sertoli cell interactions. *Mol. Endocrinol.* 3(4), 625-634.
43. Benahmed, M., Esposito, G., Sordoillet, C., dePeretti, E., Chauvin, M. A., Ghiglieri, C., Revol, A., and Morera, A. M. (1989). Transforming growth factor beta and its related peptides in the testis: an intragonadal polypeptide control system. *Persp. Androl.* 53, 191-201.
 44. Miller, D. A., Lee, A., Matsui, Y., Chen, E. Y., Moses, H. L., and Derynck, R. (1989). Complementary DNA cloning of the murine transforming growth factor-beta 3 (TGF beta 3) precursor and the comparative expression of TGF beta 3 and TGF beta 1 messenger RNA in murine embryos and adult tissues. *Mol. Endocrinol.* 3(12), 1926-1934.
 45. Mullaney, B. P., and Skinner, M. K. (1993). Transforming growth factor-beta (beta 1, beta 2, and beta 3) gene expression and action during pubertal development of the seminiferous tubule: potential role at the onset of spermatogenesis. *Mol. Endocrinol.* 7(1), 67-76.
 46. Cupp, A. S., Kim, G., and Skinner, M. K. (1999). Expression and action of transforming growth factor beta (TGFbeta1, TGFbeta2, and TGFbeta3) during embryonic rat testis development. *Biol. Reprod.* 60(6), 1304-1313.
 47. Olaso, R., Pairault, C., Saez, J. M., Habert, R., Le Magueresse-Battistoni, B., Morera, A. M., Goddard, I., and Benahmed, M. (1999). Transforming growth factor beta3 in the fetal and neonatal rat testis: immunolocalization and effect on fetal Leydig cell function. *Histochem. Cell Biol.* 112(3), 247-254.
 48. Le Magueresse-Battistoni, B., Morera, A. M., Goddard, I., and Benahmed, M. (1995). Expression of mRNAs for transforming growth factor-beta receptors in the rat testis. *Endocrinology* 136(6), 2788-2791.
 49. Caussanel, V., Tabone, E., Hendrick, J. C., Dacheux, F., and Benahmed, M. (1997). Cellular distribution of transforming growth factor betas 1, 2, and 3 and their types I and II receptors during postnatal development and spermatogenesis in the boar testis. *Biol. Reprod.* 56(2), 357-367.
 50. Olaso, R., Pairault, C., Habert, R., Caussanel, V., Tabone, E., Hendrick, J. C., Dacheux, F., and Benahmed, M. (1998). Expression of type I and II receptors for transforming growth factor beta in the adult rat testis. *Histochem. Cell Biol.* 110(6), 613-618.
 51. Ailenberg, M., Tung, P. S., and Fritz, I. B. (1990). Transforming growth factor-beta elicits shape changes and increases contractility of testicular peritubular cells. *Biol. Reprod.* 42(3), 499-509.
 52. Nargolwalla, C., McCabe, D., and Fritz, I. B. (1990). Modulation of levels of messenger RNA for tissue-type plasminogen activator in rat Sertoli cells, and levels of messenger RNA for plasminogen activator inhibitor in testis peritubular cells. *Mol. Cell Endocrinol.* 70(1), 73-80.
 53. Lui, W. Y., Lee, W. M., and Cheng, C. Y. (2001). Transforming growth factor-beta3 perturbs the inter-Sertoli tight junction permeability barrier *in vitro* possibly mediated via its effects on occludin, zonula occludens-1, and claudin-11. *Endocrinology* 142(5), 1865-1877.
 54. Lui, W. Y., Lee, W. M., and Cheng, C. Y. (2003). TGF-betas: their role in testicular function and Sertoli cell tight junction dynamics. *Int. J. Androl.* 26(3), 147-160.
 55. Avallet, O., Vigier, M., Perrard-Sapori, M. H., and Saez, J. M. (1987). Transforming growth factor beta inhibits Leydig cell functions. *Biochem. Biophys. Res. Commun.* 146(2), 575-581.
 56. Lin, T., Blaisdell, J., and Haskell, J. F. (1987). Transforming growth factor-beta inhibits Leydig cell steroidogenesis in primary culture. *Biochem. Biophys. Res. Commun.* 146(2), 387-394.
 57. Avallet, O., Vigier, M., Leduque, P., Dubois, P. M., and Saez, J. M. (1994). Expression and regulation of transforming growth factor-beta 1 messenger ribonucleic acid and protein in cultured porcine Leydig and Sertoli cells. *Endocrinology* 134(5), 2079-2087.
 58. Dobashi, M., Fujisawa, M., Yamazaki, T., Okada, H., and Kamidono, S. (2002). Distribution of intracellular and extracellular expression of transforming growth factor-beta1 (TGF-beta1) in human testis and their association with spermatogenesis. *Asian J. Androl.* 4(2), 105-109.
 59. Haagmans, B. L., Hoogerbrugge, J. W., Themmen, A. P., and Teerds, K. J. (2003). Rat testicular germ cells and Sertoli cells release different types of bioactive transforming growth factor beta *in vitro*. *Reprod. Biol. Endocrinol.* 1(1), 3.
 60. Loveland, K. L., Bakker, M., Meehan, T., Christy, E., von Schonfeldt, V., Drummond, A., de Kretser, D., Haagmans, B. L., Hoogerbrugge, J. W., Themmen, A. P., and Teerds, K. J. (2003). Expression of Bambi is widespread in juvenile and adult rat tissues and is regulated in male germ cells. *Endocrinology* 144(9), 4180-4186.
 61. Gospodarowicz, D., and Ferrara, N. (1989). Fibroblast growth factor and the control of pituitary and gonad development and function. *J. Steroid Biochem.* 32(1B), 183-191.
 62. Ueno, N., Ling, N., Ying, S. Y., Esch, F., Shimasaki, S., and Guillemin, R. (1987). Isolation and partial characterization of follistatin: a single-chain Mr 35,000 monomeric protein that inhibits the release of follicle-stimulating hormone. *Proc. Natl. Acad. Sci. USA* 84(23), 8282-8286.
 63. Story, M. T., Sasse, J., Kakuska, D., Jacobs, S. C., and Lawson, R. K. (1988). A growth factor in bovine and human testes structurally related to basic fibroblast growth factor. *J. Urol.* 140(2), 422-427.
 64. Smith, E. P., Hall, S. H., Monaco, L., French, F. S., Wilson, E. M., and Conti, M. (1989). A rat Sertoli cell factor similar to basic fibroblast growth factor increases c-fos messenger ribonucleic acid in cultured Sertoli cells. *Mol. Endocrinol.* 3(6), 954-961.
 65. Mullaney, B. P., and Skinner, M. K. (1992). Basic fibroblast growth factor (bFGF) gene expression and protein production during pubertal development of the seminiferous tubule: follicle-stimulating hormone-induced Sertoli cell bFGF expression. *Endocrinology* 131(6), 2928-2934.
 66. Jaillard, C., Chatelain, P. G., and Saez, J. M. (1987). *In vitro* regulation of pig Sertoli cell growth and function: effects of fibroblast growth factor and somatomedin-C. *Biol. Reprod.* 37(3), 665-674.
 67. Van Dissel-Emiliani, F. M., De Boer-Brouwer, M., and De Rooij, D. G. (1996). Effect of fibroblast growth factor-2 on Sertoli cells and gonocytes in coculture during the perinatal period. *Endocrinology* 137(2), 647-654.
 68. Brucato, S., Bocquet, J., and Villers, C. (2002). Cell surface heparan sulfate proteoglycans: target and partners of the basic fibroblast growth factor in rat Sertoli cells. *Eur. J. Biochem.* 269(2), 502-511.
 69. Riera, M. F., Meroni, S. B., Schteingart, H. F., Pellizzari, E. H., and Cigorraga, S. B. (2002). Regulation of lactate production and glucose transport as well as of glucose transporter 1 and lactate dehydrogenase A mRNA levels by basic fibroblast growth factor in rat Sertoli cells. *J. Endocrinol.* 173(2), 335-343.
 70. Le Magueresse-Battistoni, B., Wolff, J., Morera, A. M., and Benahmed, M. (1994). Fibroblast growth factor receptor type 1 expression during rat testicular development and its regulation in cultured Sertoli cells. *Endocrinology* 135(6), 2404-2411.
 71. Yamamoto, H., Ochiya, T., Takahama, Y., Ishii, Y., Osumi, N., Sakamoto, H., and Terada, M. (2000). Detection of spatial localization of Hst-1/Fgf-4 gene expression in brain and testis from adult mice. *Oncogene* 19(33), 3805-3810.
 72. Yamamoto, H., Ochiya, T., Tamamushi, S., Toriyama-Baba, H., Takahama, Y., Hirai, K., Sasaki, H., Sakamoto, H., Saito, I., Iwamoto, T., Kakizoe, T., and Terada, M. (2002). HST-1/FGF-4 gene activation induces spermatogenesis and prevents adriamycin-induced testicular toxicity. *Oncogene* 21(6), 899-908.

73. Colvin, J. S., Green, R. P., Schmahl, J., Capel, B., and Ornitz, D. M. (2001). Male-to-female sex reversal in mice lacking fibroblast growth factor 9. *Cell* **104**(6), 875–889.
74. Olson, L., Ayer-LeLievre, C., Ebendal, T., and Seiger, A. (1987). Nerve growth factor-like immunoreactivities in rodent salivary glands and testis. *Cell Tissue Res.* **248**(2), 275–286.
75. Ayer-LeLievre, C., Olson, L., Ebendal, T., Hallbook, F., and Persson, H. (1988). Nerve growth factor mRNA and protein in the testis and epididymis of mouse and rat. *Proc. Natl. Acad. Sci. USA* **85**(8), 2628–2632.
76. Chen, Y., Dicou, E., and Djakiew, D. (1997). Characterization of nerve growth factor precursor protein expression in rat round spermatids and the trophic effects of nerve growth factor in the maintenance of Sertoli cell viability. *Mol. Cell Endocrinol.* **127**(2), 129–136.
77. Persson, H., Ayer-LeLievre, C., Soder, O., Villar, M. J., Metsis, M., Olson, L., Ritzen, M., and Hokfelt, T. (1990). Expression of beta-nerve growth factor receptor mRNA in Sertoli cells down-regulated by testosterone. *Science* **247**(4943), 704–707.
78. Scheingart, H. F., Meroni, S. B., Canepa, D. F., Pellizzari, E. H., and Cigorraga, S. B. (1999). Effects of basic fibroblast growth factor and nerve growth factor on lactate production, gamma-glutamyl transpeptidase and aromatase activities in cultured Sertoli cells. *Eur. J. Endocrinol.* **141**(5), 539–545.
79. Seidl, K., Buchberger, A., and Erck, C. (1996). Expression of nerve growth factor and neurotrophin receptors in testicular cells suggest novel roles for neurotrophins outside the nervous system. *Reprod. Fertil. Dev.* **8**(7), 1075–1087.
80. Djakiew, D., Pflug, B., Dionne, C., and Onoda, M. (1994). Postnatal expression of nerve growth factor receptors in the rat testis. *Biol. Reprod.* **51**(2), 214–221.
81. Cupp, A. S., Kim, G. H., and Skinner, M. K. (2000). Expression and action of neurotrophin-3 and nerve growth factor in embryonic and early postnatal rat testis development. *Biol. Reprod.* **63**(6), 1617–1628.
82. Park, C., Choi, W. S., Kwon, H., and Kwon, Y. K. (2001). Temporal and spatial expression of neurotrophins and their receptors during male germ cell development. *Mol. Cells* **12**(3), 360–367.
83. Levine, E., Cupp, A. S., and Skinner, M. K. (2000). Role of neurotrophins in rat embryonic testis morphogenesis (cord formation). *Biol. Reprod.* **62**(1), 132–142.
84. Cupp, A. S., Uzumcu, M., and Skinner, M. K. (2003). Chemotactic role of neurotrophin 3 in the embryonic testis that facilitates male sex determination. *Biol. Reprod.* **68**(6), 2033–2037.
85. Robinson, L. L., Townsend, J., and Anderson, R. A. (2003). The human fetal testis is a site of expression of neurotrophins and their receptors: regulation of the germ cell and peritubular cell population. *J. Clin. Endocrinol. Metab.* **88**(8), 3943–3951.
86. Syed, V., Soder, O., Arver, S., Lindh, M., Khan, S., and Ritzen, E. M. (1988). Ontogeny and cellular origin of an interleukin-1-like factor in the reproductive tract of the male rat. *Int. J. Androl.* **11**(5), 437–447.
87. Pollanen, P., Soder, O., and Parvinen, M. (1989). Interleukin-1 alpha stimulation of spermatogonial proliferation *in vivo*. *Reprod. Fertil. Dev.* **1**(1), 85–87.
88. Jonsson, C. K., Zetterstrom, R. H., Holst, M., Parvinen, M., and Soder, O. (1999). Constitutive expression of interleukin-1alpha messenger ribonucleic acid in rat Sertoli cells is dependent upon interaction with germ cells. *Endocrinology* **140**(8), 3755–3761.
89. Petersen, C., Boitani, C., Froysa, B., and Soder, O. (2002). Interleukin-1 is a potent growth factor for immature rat Sertoli cells. *Mol. Cell Endocrinol.* **186**(1), 37–47.
90. Huleihel, M., and Lunenfeld, E. (2002). Involvement of intratesticular IL-1 system in the regulation of Sertoli cell functions. *Mol. Cell Endocrinol.* **187**(1–2), 125–132.
91. Nehar, D., Mauduit, C., Boussouar, F., and Benahmed, M. (1998). Interleukin 1alpha stimulates lactate dehydrogenase A expression and lactate production in cultured porcine Sertoli cells. *Biol. Reprod.* **59**(6), 1425–1432.
92. Zeyse, D., Lunenfeld, E., Beck, M., Prinsloo, I., and Huleihel, M. (2000). Interleukin-1 receptor antagonist is produced by Sertoli cells *in vitro*. *Endocrinology* **141**(4), 1521–1527.
93. Huleihel, M., Zeyse, D., Lunenfeld, E., Beck, M., Prinsloo, I., Potashnik, G., and Mazor, M. (2001). Immunohistochemical staining of IL-1 alpha and IL-1 receptor antagonist but not IL-1 beta in cultures of Sertoli cells. *Am. J. Reprod. Immunol.* **45**(3), 135–141.
94. Soder, O., Sultana, T., Jonsson, C., Wahlgren, A., Petersen, C., and Holst, M. (2000). The interleukin-1 system in the testis. *Andrologia* **32**(1), 52–55.
95. Zeyse, D., Lunenfeld, E., Beck, M., Prinsloo, I., and Huleihel, M. (2000). Induction of interleukin-1alpha production in murine Sertoli cells by interleukin-1. *Biol. Reprod.* **62**(5), 1291–1296.
96. Syed, V., Stephan, J. P., Gerard, N., Legrand, A., Parvinen, M., Bardin, C. W., and Jegou, B. (1995). Residual bodies activate Sertoli cell interleukin-1 alpha (IL-1 alpha) release, which triggers IL-6 production by an autocrine mechanism, through the lipoxygenase pathway. *Endocrinology* **136**(7), 3070–3078.
97. Okuda, Y., Bardin, C. W., Hodgskin, L. R., and Morris, P. L. (1995). Interleukins-1 alpha and -1 beta regulate interleukin-6 expression in Leydig and Sertoli cells. *Recent Prog. Horm. Res.* **50**, 367–372.
98. Jegou, B., Cudicini, C., Gomez, E., and Stephan, J. P. (1995). Interleukin-1, interleukin-6 and the germ cell-Sertoli cell cross-talk. *Reprod. Fertil. Dev.* **7**(4), 723–730.
99. Stephan, J. P., Syed, V., and Jegou, B. (1997). Regulation of Sertoli cell IL-1 and IL-6 production *in vitro*. *Mol. Cell Endocrinol.* **134**(2), 109–118.
100. Cudicini, C., Kercret, H., Touzalin, A. M., Ballet, F., and Jegou, B. (1997). Vectorial production of interleukin 1 and interleukin 6 by rat Sertoli cells cultured in a dual culture compartment system. *Endocrinology* **138**(7), 2863–2870.
101. Jenab, S., and Morris, P. L. (2000). Interleukin-6 regulation of kappa opioid receptor gene expression in primary Sertoli cells. *Endocrine* **13**(1), 11–15.
102. Fujisawa, M., Okuda, Y., Fujioka, H., and Kamidono, S. (2002). Expression and regulation of gp130 messenger ribonucleic acid in cultured immature rat Sertoli cells. *Endocr. Res.* **28**(1–2), 1–8.
103. Hoeben, E., Wuyts, A., Proost, P., Van Damme, J., and Verhoeven, G. (1997). Identification of IL-6 as one of the important cytokines responsible for the ability of mononuclear cells to stimulate Sertoli cell functions. *Mol. Cell Endocrinol.* **132**(1–2), 149–160.
104. Riccioli, A., Filippini, A., De Cesaris, P., Barbacci, E., Stefanini, M., Starace, G., and Ziparo, E. (1995). Inflammatory mediators increase surface expression of integrin ligands, adhesion to lymphocytes, and secretion of interleukin 6 in mouse Sertoli cells. *Proc. Natl. Acad. Sci. USA* **92**(13), 5808–5812.
105. Sultana, T., Wahab-Wahlgren, A., Assmus, M., Parvinen, M., Weber, G., and Soder, O. (2003). Expression and regulation of the prointerleukin-1alpha processing enzymes calpain I and II in the rat testis. *Int. J. Androl.* **26**(1), 37–45.
106. Cudicini, C., Lejeune, H., Gomez, E., Bosmans, E., Ballet, F., Saez, J., and Jegou, B. (1997). Human Leydig cells and Sertoli cells are producers of interleukins-1 and -6. *J. Clin. Endocrinol. Metab.* **82**(5), 1426–1433.
107. Dejuq, N., Dugast, I., Ruffault, A., van der Meide, P. H., and Jegou, B. (1995). Interferon-alpha and -gamma expression in the rat testis. *Endocrinology* **136**(11), 4925–4931.
108. Braghirioli, L., Silvestrini, B., Sorrentino, C., Grima, J., Mruk, D., and Cheng, C. Y. (1998). Regulation of alpha2-macroglobulin

- expression in rat Sertoli cells and hepatocytes by germ cells *in vitro*. *Biol. Reprod.* **59**(1), 111–123.
109. Meinhardt, A., Bacher, M., Wennemuth, G., Eickhoff, R., and Hedger, M. (2000). Macrophage migration inhibitory factor (MIF) as a paracrine mediator in the interaction of testicular somatic cells. *Andrologia* **32**(1), 46–48.
 110. Wennemuth, G., Aumuller, G., Bacher, M., and Meinhardt, A. (2000). Macrophage migration inhibitory factor-induced Ca(2+) response in rat testicular peritubular cells. *Biol. Reprod.* **62**(6), 1632–1639.
 111. Aubry, F., Habasque, C., Satie, A. P., Jegou, B., and Samson, M. (2000). Expression and regulation of the CC-chemokine monocyte chemoattractant protein-1 in rat testicular cells in primary culture. *Biol. Reprod.* **62**(5), 1427–1435.
 112. Vincent, S., Segretain, D., Nishikawa, S., Nishikawa, S. I., Sage, J., Cuzin, F., and Rassoulzadegan, M. (1998). Stage-specific expression of the Kit receptor and its ligand (KL) during male gametogenesis in the mouse: a Kit-KL interaction critical for meiosis. *Development*. **125**(22), 4585–4593.
 113. Hakovirta, H., Yan, W., Kaleva, M., Zhang, F., Vanttinen, K., Morris, P. L., Soder, M., Parvinen, M., and Toppari, J. (1999). Function of stem cell factor as a survival factor of spermatogonia and localization of messenger ribonucleic acid in the rat seminiferous epithelium. *Endocrinology* **140**(3), 1492–1498.
 114. Orth, J. M., Qiu, J., Jester, W. F., Jr., and Pilder, S. (1997). Expression of the c-kit gene is critical for migration of neonatal rat gonocytes *in vitro*. *Biol. Reprod.* **57**(3), 676–683.
 115. Blanchard, K. T., Lee, J., and Boekelheide, K. (1998). Leuprolide, a gonadotropin-releasing hormone agonist, reestablishes spermatogenesis after 2,5-hexanedione-induced irreversible testicular injury in the rat, resulting in normalized stem cell factor expression. *Endocrinology* **139**(1), 236–244.
 116. Steinmetz, R., Lazzaro, N., Rothrock, J. K., and Pescovitz, O. H. (2000). Effects of growth hormone-releasing hormone-related peptide on stem cell factor expression in cultured rat Sertoli cells. *Endocrine* **12**(3), 323–327.
 117. Goddard, I., Bauer, S., Gougeon, A., Lopez, F., Giannetti, N., Susini, C., Benahmed, M., and Krantic, S. (2001). Somatostatin inhibits stem cell factor messenger RNA expression by Sertoli cells and stem cell factor-induced DNA synthesis in isolated seminiferous tubules. *Biol. Reprod.* **65**(6), 1732–1742.
 118. Feng, H. L., Sandlow, J. I., Sparks, A. E., Sandra, A., and Zheng, L. J. (1999). Decreased expression of the c-kit receptor is associated with increased apoptosis in subfertile human testes. *Fertil. Steril.* **71**(1), 85–89.
 119. Grimaldi, P., Rossi, P., Dolci, S., Ripamonti, C. B., and Geremia, R. (2002). Molecular genetics of male infertility: stem cell factor/c-kit system. *Am. J. Reprod. Immunol.* **48**(1), 27–33.
 120. Munsie, M., Schlatt, S., deKretser, D. M., and Loveland, K. L. (1997). Expression of stem cell factor in the postnatal rat testis. *Mol. Reprod. Dev.* **47**(1), 19–25.
 121. Yan, W., Kero, J., Huhtaniemi, I., and Toppari, J. (2000). Stem cell factor functions as a survival factor for mature Leydig cells and a growth factor for precursor Leydig cells after ethylene dimethane sulfonate treatment: implication of a role of the stem cell factor/c-Kit system in Leydig cell development. *Dev. Biol.* **227**(1), 169–182.
 122. Meng, X., Lindahl, M., Hyvonen, M. E., Parvinen, M., de Rooij, D. G., Hess, M. W., Raatikainen-Ahokas, A., Sainio, K., Rauvala, H., Lakso, M., Pichel, J. G., Westphal, H., Saarma, M., and Sariola, H. (2000). Regulation of cell fate decision of undifferentiated spermatogonia by GDNF. *Science* **287**(5457), 1489–1493.
 123. Davidoff, M. S., Middendorff, R., Koeva, Y., Pusch, W., Jezek, D., and Muller, D. (2001). Glial cell line-derived neurotrophic factor (GDNF) and its receptors GFRalpha-1 and GFRalpha-2 in the human testis. *Ital. J. Anat. Embryol.* **106**(2 Suppl 2), 173–180.
 124. Tadokoro, Y., Yomogida, K., Ohta, H., Tohda, A., and Nishimune, Y. (2002). Homeostatic regulation of germinal stem cell proliferation by the GDNF/FSH pathway. *Mech. Dev.* **113**(1), 29–39.
 125. Viglietto, G., Dolci, S., Bruni, P., Baldassarre, G., Chiariotti, L., Melillo, R. M., Salvatore, G., Chiappetta, G., Sferratore, F., Fusco, A., and Santoro, M. (2000). Glial cell line-derived neurotrophic factor and neurturin can act as paracrine growth factors stimulating DNA synthesis of Ret-expressing spermatogonia. *Int. J. Oncol.* **16**(4), 689–694.
 126. Meng, X., Pata, I., Pedrono, E., Popsueva, A., de Rooij, D. G., Janne, M., Rauvala, H., and Sariola, H. (2001). Transient disruption of spermatogenesis by deregulated expression of neurturin in testis. *Mol. Cell Endocrinol.* **184**(1–2), 33–39.
 127. Sariola, H., and Meng, X. (2003). GDNF-induced seminomatous tumours in mouse—an experimental model for human seminomas? *Apmis* **111**(1), 192–196; discussion 196.
 128. Hu, J., Shima, H., and Nakagawa, H. (1999). Glial cell line-derived neurotrophic factor stimulates Sertoli cell proliferation in the early postnatal period of rat testis development. *Endocrinology* **140**(8), 3416–3421.
 129. McCullagh, D. R. (1932). Dual endocrine activity of the testis. *Science* **76**, 19–20.
 130. Rivier, C., Vale, W., and Rivier, J. (1987). Studies of the inhibin family of hormones: a review. *Horm. Res.* **28**(2–4), 104–118.
 131. Risbridger, G. P., Robertson, D. M., and de Kretser, D. M. (1990). Current perspectives of inhibin biology. *Acta Endocrinol. (Copenhagen)* **122**(6), 673–682.
 132. Steinberger, A., and Steinberger, E. (1976). Secretion of an FSH-inhibiting factor by cultured Sertoli cells. *Endocrinology* **99**(3), 918–921.
 133. Bicsak, T. A., Vale, W., Vaughan, J., Tucker, E. M., Cappel, S., and Hsueh, A. J. (1987). Hormonal regulation of inhibin production by cultured Sertoli cells. *Mol. Cell Endocrinol.* **49**(2–3), 211–217.
 134. Morris, P. L., Vale, W. W., Cappel, S., and Bardin, C. W. (1988). Inhibin production by primary Sertoli cell-enriched cultures: regulation by follicle-stimulating hormone, androgens, and epidermal growth factor. *Endocrinology* **122**(2), 717–725.
 135. Gonzales, G. F., Risbridger, G. P., and de Kretser, D. M. (1988). *In vitro* synthesis and release of inhibin in response to FSH stimulation by isolated segments of seminiferous tubules from normal adult male rats. *Mol. Cell Endocrinol.* **59**(3), 179–185.
 136. Meehan, T., Schlatt, S., O'Bryan, M. K., de Kretser, D. M., and Loveland, K. L. (2000). Regulation of germ cell and Sertoli cell development by activin, follistatin, and FSH. *Dev. Biol.* **220**(2), 225–237.
 137. de Kretser, D. M., Meinhardt, A., Meehan, T., Phillips, D. J., O'Bryan, M. K., and Loveland, K. A. (2000). The roles of inhibin and related peptides in gonadal function. *Mol. Cell Endocrinol.* **161**(1–2), 43–46.
 138. Boitani, C., Stefanini, M., Fragale, A., and Morena, A. R. (1995). Activin stimulates Sertoli cell proliferation in a defined period of rat testis development. *Endocrinology* **136**(12), 5438–5444.
 139. van Dissel-Emiliani, F. M., Grootenhuys, A. J., de Jong, F. H., and de Rooij, D. G. (1989). Inhibin reduces spermatogonial numbers in testes of adult mice and Chinese hamsters. *Endocrinology* **125**(4), 1899–1903.
 140. Marchetti, C., Hamdane, M., Mitchell, V., Mayo, K., Devisme, L., Rigot, J. M., Beauvillain, J. C., Hermand, E., and Defossez, A. (2003). Immunolocalization of inhibin and activin alpha and betaB subunits and expression of corresponding messenger RNAs in the human adult testis. *Biol. Reprod.* **68**(1), 230–235.

141. Clifton, R. J., O'Donnell, L., and Robertson, D. M. (2002). Pachytene spermatocytes in co-culture inhibit rat Sertoli cell synthesis of inhibin beta B-subunit and inhibin B but not the inhibin alpha-subunit. *J. Endocrinol.* **172**(3), 565–574.
142. Risbridger, G. P., Clements, J., Robertson, D. M., Drummond, A. E., Muir, J., Burger, H. G., and de Kretser, D. M. (1989). Immunological and bioactive inhibin and inhibin alpha-subunit expression in rat Leydig cell cultures. *Mol. Cell Endocrinol.* **66**(1), 119–122.
143. Roberts, V., Meunier, H., Sawchenko, P. E., and Vale, W. (1989). Differential production and regulation of inhibin subunits in rat testicular cell types. *Endocrinology* **125**(5), 2350–2359.
144. Shaha, C., Morris, P. L., Chen, C. L., Vale, W., and Bardin, C. W. (1989). Immunostainable inhibin subunits are in multiple types of testicular cells. *Endocrinology* **125**(4), 1941–1950.
145. Lee, W., Mason, A. J., Schwall, R., Szonyi, E., and Mather, J. P. (1989). Secretion of activin by interstitial cells in the testis. *Science* **243**(4889), 396–398.
146. de Winter, J. P., Vanderstichele, H. M., Verhoeven, G., Timmerman, M. A., Wesseling, J. G., and de Jong, F. H. (1994). Peritubular myoid cells from immature rat testes secrete activin-A and express activin receptor type II *in vitro*. *Endocrinology* **135**(2), 759–767.
147. Buzzard, J. J., Farnworth, P. G., De Kretser, D. M., O'Connor, A. E., Wreford, N. G., and Morrison, J. R. (2003). Proliferative phase Sertoli cells display a developmentally regulated response to activin *in vitro*. *Endocrinology* **144**(2), 474–483.
148. Anawalt, B. D., Bebb, R. A., Matsumoto, A. M., Groome, N. P., Illingworth, P. J., McNeilly, A. S., and Bremner, W. J. (1996). Serum inhibin B levels reflect Sertoli cell function in normal men and men with testicular dysfunction. *J. Clin. Endocrinol. Metab.* **81**(9), 3341–3345.
149. Crofton, P. M., Evans, A. E., Groome, N. P., Taylor, M. R., Holland, C. V., and Kelnar, C. J. (2002). Inhibin B in boys from birth to adulthood: relationship with age, pubertal stage, FSH and testosterone. *Clin. Endocrinol. (Oxford)* **56**(2), 215–221.
150. Crofton, P. M., Thomson, A. B., Evans, A. E., Groome, N. P., Bath, L. E., Kelnar, C. J., and Wallace, W. H. (2003). Is inhibin B a potential marker of gonadotoxicity in prepubertal children treated for cancer? *Clin. Endocrinol. (Oxford)* **58**(3), 296–301.
151. Josso, N., di Clemente, N., and Gouedard, L. (2001). Anti-Müllerian hormone and its receptors. *Mol. Cell Endocrinol.* **179**(1–2), 25–32.
152. Behringer, R. R. (1995). The Müllerian inhibitor and mammalian sexual development. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* **350**(1333), 285–288; discussion 289.
153. Jost, A. (1953). Problems of fetal endocrinology: the gonadal and hypophyseal hormones. *Recent Prog. Horm. Res* **8**, 379–418.
154. Hayashi, M., Shima, H., Hayashi, K., Trelstad, R. L., and Donahoe, P. K. (1984). Immunocytochemical localization of Müllerian inhibiting substance in the rough endoplasmic reticulum and Golgi apparatus in Sertoli cells of the neonatal calf testis using a monoclonal antibody. *J. Histochem. Cytochem.* **32**(6), 649–654.
155. Blanchard, M. G., and Josso, N. (1974). Source of the anti-Müllerian hormone synthesized by the fetal testis: Müllerian-inhibiting activity of fetal bovine Sertoli cells in tissue culture. *Pediatr. Res.* **8**(12), 968–971.
156. Cate, R. L., Mattaliano, R. J., Hession, C., Tizard, R., Farber, N. M., Cheung, A., Ninfa, E. G., Frey, A. Z., Gash, D. J., Chow, E. P., et al. (1986). Isolation of the bovine and human genes for Müllerian inhibiting substance and expression of the human gene in animal cells. *Cell* **45**(5), 685–698.
157. Hutson, J., Ikawa, H., and Donahoe, P. K. (1981). The ontogeny of Müllerian inhibiting substance in the gonads of the chicken. *J. Pediatr. Surg.* **16**(6), 822–827.
158. Juengel, J. L., Whale, L. J., Wylde, K. A., Greenwood, P., McNatty, K. P., and Eckery, D. C. (2002). Expression of anti-Müllerian hormone mRNA during gonadal and follicular development in the brushtail possum (*Trichosurus vulpecula*). *Reprod. Fertil. Dev.* **14**(5–6), 345–353.
159. Kuroda, T., Lee, M. M., Haqq, C. M., Powell, D. M., Manganaro, T. F., and Donahoe, P. K. (1990). Müllerian inhibiting substance ontogeny and its modulation by follicle-stimulating hormone in the rat testes. *Endocrinology* **127**(4), 1825–1832.
160. Teixeira, J., Fynn-Thompson, E., Payne, A. H., and Donahoe, P. K. (1999). Müllerian-inhibiting substance regulates androgen synthesis at the transcriptional level. *Endocrinology* **140**(10), 4732–4738.
161. Watanabe, K., Clarke, T. R., Lane, A. H., Wang, X., and Donahoe, P. K. (2000). Endogenous expression of Müllerian inhibiting substance in early postnatal rat Sertoli cells requires multiple steroidogenic factor-1 and GATA-4-binding sites. *Proc. Natl. Acad. Sci. USA* **97**(4), 1624–1629.
162. Tevosian, S. G., Albrecht, K. H., Crispino, J. D., Fujiwara, Y., Eicher, E. M., and Orkin, S. H. (2002). Gonadal differentiation, sex determination and normal Sry expression in mice require direct interaction between transcription partners GATA4 and FOG2. *Development* **129**(19), 4627–4634.
163. Schepers, G., Wilson, M., Wilhelm, D., and Koopman, P. (2003). SOX8 is expressed during testis differentiation in mice and synergizes with SF1 to activate the AMH promoter *in vitro*. *J. Biol. Chem.* **278**(30), 28101–28108.
164. Al-Attar, L., Noel, K., Dutertre, M., Belville, C., Forest, M. G., Burgoyne, P. S., Josso, N., and Rey, R. (1997). Hormonal and cellular regulation of Sertoli cell anti-Müllerian hormone production in the postnatal mouse. *J. Clin. Invest.* **100**(6), 1335–1343.
165. Arambepola, N. K., Bunick, D., and Cooke, P. S. (1998). Thyroid hormone and follicle-stimulating hormone regulate Müllerian-inhibiting substance messenger ribonucleic acid expression in cultured neonatal rat Sertoli cells. *Endocrinology* **139**(11), 4489–4495.
166. Blagosklonova, O., Joanne, C., Roux, C., Bittard, H., Fellmann, F., and Bresson, J. L. (2002). Absence of anti-Müllerian hormone (AMH) and M2A immunoreactivities in Sertoli cell-only syndrome and maturation arrest with and without AZF microdeletions. *Hum. Reprod.* **17**(8), 2062–2065.
167. Lang-Muritano, M., Biason-Lauber, A., Gitzelmann, C., Belville, C., Picard, Y., and Schoenle, E. J. (2001). A novel mutation in the anti-Müllerian hormone gene as cause of persistent Müllerian duct syndrome. *Eur. J. Pediatr.* **160**(11), 652–654.
168. Fenichel, P., Rey, R., Poggioli, S., Donzeau, M., Chevallier, D., and Pointis, G. (1999). Anti-Müllerian hormone as a seminal marker for spermatogenesis in non-obstructive azoospermia. *Hum. Reprod.* **14**(8), 2020–2024.
169. Eddy, E. M., Washburn, T. F., Bunch, D. O., Goulding, E. H., Gladen, B. C., Lubahn, D. B., and Korach, K. S. (1996). Targeted disruption of the estrogen receptor gene in male mice causes alteration of spermatogenesis and infertility. *Endocrinology* **137**(11), 4796–4805.
170. Lee, K. H., Hess, R. A., Bahr, J. M., Lubahn, D. B., Taylor, J., and Bunick, D. (2000). Estrogen receptor alpha has a functional role in the mouse rete testis and efferent ductules. *Biol. Reprod.* **63**(6), 1873–1880.
171. Nakai, M., Bouma, J., Nie, R., Zhou, Q., Carnes, K., Lubahn, D. B., and Hess, R. A. (2001). Morphological analysis of endocytosis in efferent ductules of estrogen receptor-alpha knockout male mouse. *Anat. Rec.* **263**(1), 10–18.

172. O'Donnell, L., Robertson, K. M., Jones, M. E., and Simpson, E. R. (2001). Estrogen and spermatogenesis. *Endocr. Rev.* **22**(3), 289–318.
173. Pelletier, G., Labrie, C., and Labrie, F. (2000). Localization of oestrogen receptor alpha, oestrogen receptor beta and androgen receptors in the rat reproductive organs. *J. Endocrinol.* **165**(2), 359–370.
174. Saunders, P. T., Fisher, J. S., Sharpe, R. M., and Millar, M. R. (1998). Expression of oestrogen receptor beta (ER beta) occurs in multiple cell types, including some germ cells, in the rat testis. *J. Endocrinol.* **156**(3), R13–17.
175. MacCalman, C. D., Getsios, S., Farookhi, R., and Blaschuk, O. W. (1997). Estrogens potentiate the stimulatory effects of follicle-stimulating hormone on N-cadherin messenger ribonucleic acid levels in cultured mouse Sertoli cells. *Endocrinology* **138**(1), 41–48.
176. Feig, L. A., Bellve, A. R., Erickson, N. H., and Klagsbrun, M. (1980). Sertoli cells contain a mitogenic polypeptide. *Proc. Natl. Acad. Sci. USA* **77**(8), 4774–4778.
177. Feig, L. A., Klagsbrun, M., and Bellve, A. R. (1983). Mitogenic polypeptide of the mammalian seminiferous epithelium: biochemical characterization and partial purification. *J. Cell Biol.* **97**(5 Pt 1), 1435–1443.
178. Bellve, A. R., and Feig, L. A. (1984). Cell proliferation in the mammalian testis: biology of the seminiferous growth factor (SGF). *Recent Prog. Horm. Res.* **40**, 531–567.
179. Buch, J. P., Lamb, D. J., Lipshultz, L. I., and Smith, R. G. (1988). Partial characterization of a unique growth factor secreted by human Sertoli cells. *Fertil. Steril.* **49**(4), 658–665.
180. Rich, K. A., and De Kretser, D. M. (1977). Effect of differing degrees of destruction of the rat seminiferous epithelium on levels of serum follicle stimulating hormone and androgen binding protein. *Endocrinology* **101**(3), 959–968.
181. Aoki, A., and Fawcett, D. W. (1978). Is there a local feedback from the seminiferous tubules affecting activity of the Leydig cells? *Biol. Reprod.* **19**(1), 144–158.
182. Rich, K. A., Kerr, J. B., and de Kretser, D. M. (1979). Evidence for Leydig cell dysfunction in rats with seminiferous tubule damage. *Mol. Cell Endocrinol.* **13**(2), 123–135.
183. de Kretser, D. M., Sharpe, R. M., and Swanston, I. A. (1979). Alterations in steroidogenesis and human chorionic gonadotropin binding in the cryptorchid rat testis. *Endocrinology* **105**(1), 135–138.
184. Risbridger, G. P., Kerr, J. B., Peake, R., Rich, K. A., and de Kretser, D. M. (1981). Temporal changes in rat Leydig cell function after the induction of bilateral cryptorchidism. *J. Reprod. Fertil.* **63**(2), 415–423.
185. Bergh, A. (1983). Paracrine regulation of Leydig cells by the seminiferous tubules. *Int. J. Androl.* **6**(1), 57–65.
186. Verhoeven, G., and Cailleau, J. (1985). A factor in spent media from Sertoli-cell-enriched cultures that stimulates steroidogenesis in Leydig cells. *Mol. Cell Endocrinol.* **40**(1), 57–68.
187. Benahmed, M., Tabone, E., Grenot, C., Sanchez, P., Chauvin, M. A., and Morera, A. M. (1986). Paracrine control of Leydig cell activity by FSH dependent proteins from Sertoli cells: an *in vitro* study. *J. Steroid Biochem.* **24**(1), 311–315.
188. Verhoeven, G., and Cailleau, J. (1986). Specificity and partial purification of a factor in spent media from Sertoli cell-enriched cultures that stimulates steroidogenesis in Leydig cells. *J. Steroid Biochem.* **25**(3), 393–402.
189. Carreau, S., Papadopoulos, V., and Drosowsky, M. A. (1988). Stimulation of adult rat Leydig cell aromatase activity by a Sertoli cell factor. *Endocrinology* **122**(3), 1103–1109.
190. Papadopoulos, V., Kamtchouing, P., Drosowsky, M. A., and Carreau, S. (1987). Spent media from immature seminiferous tubules and Sertoli cells inhibit adult rat Leydig cell aromatase activity. *Horm. Metab. Res.* **19**(2), 62–64.
191. Syed, V., Khan, S. A., Lindh, M., and Ritzen, E. M. (1987). Ontogeny and cellular origin of a rat seminiferous tubule factor(s) that inhibits LH-dependent testosterone production by interstitial cells *in vitro*. *Int. J. Androl.* **10**(5), 711–720.
192. Vihko, K. K., and Huhtaniemi, I. (1989). A rat seminiferous epithelial factor that inhibits Leydig cell cAMP and testosterone production: mechanism of action, stage-specific secretion, and partial characterization. *Mol. Cell Endocrinol.* **65**(1–2), 119–127.
193. Wu, N., and Muroso, E. P. (1994). A Sertoli cell-secreted paracrine factor(s) stimulates proliferation and inhibits steroidogenesis of rat Leydig cells. *Mol. Cell Endocrinol.* **106**(1–2), 99–109.
194. Magnanti, M., Gandini, O., Giuliani, L., Gazzaniga, P., Marti, H. H., Gradilone, A., Frati, L., Agliano, A. M., and Gassmann, M. (2001). Erythropoietin expression in primary rat Sertoli and peritubular myoid cells. *Blood* **98**(9), 2872–2874.
195. De Miguel, M. P., De Boer-Brouwer, M., Paniagua, R., van den Hurk, R., De Rooij, D. G., and Van Dissel-Emiliani, F. M. (1996). Leukemia inhibitory factor and ciliary neurotropic factor promote the survival of Sertoli cells and gonocytes in coculture system. *Endocrinology* **137**(5), 1885–1893.
196. McLean, D. J., Friel, P. J., Pouchnik, D., and Griswold, M. D. (2002). Oligonucleotide microarray analysis of gene expression in follicle-stimulating hormone-treated rat Sertoli cells. *Mol. Endocrinol.* **16**(12), 2780–2792.
197. Kranenburg, O., and Moolenaar, W. H. (2001). Ras-MAP kinase signaling by lysophosphatidic acid and other G protein-coupled receptor agonists. *Oncogene* **20**(13), 1540–1546.
198. Chun, J., Contos, J. J., and Munroe, D. (1999). A growing family of receptor genes for lysophosphatidic acid (LPA) and other lysophospholipids (LPs). *Cell Biochem. Biophys.* **30**(2), 213–242.
199. Sonoda, H., Aoki, J., Hiramatsu, T., Ishida, M., Bandoh, K., Nagai, Y., Taguchi, R., Inoue, K., and Arai, H. (2002). A novel phosphatidic acid-selective phospholipase A1 that produces lysophosphatidic acid. *J. Biol. Chem.* **277**(37), 34254–34263.
200. Riffo, M. S., and Parraga, M. (1996). Study of the acrosome reaction and the fertilizing ability of hamster epididymal cauda spermatozoa treated with antibodies against phospholipase A2 and/or lysophosphatidylcholine. *J. Exp. Zool.* **275**(6), 459–468.
201. Therien, I., Bleau, G., and Manjunath, P. (1995). Phosphatidylcholine-binding proteins of bovine seminal plasma modulate capacitation of spermatozoa by heparin. *Biol. Reprod.* **52**(6), 1372–1379.
202. Meroni, S. B., Canepa, D. F., Pellizzari, E. H., Schteingart, H. F., and Cigorraga, S. B. (1999). Effect of N-acetylsphingosine (C2) and the ceramidase inhibitor (1S,2R)-D-erythro-2-(N-myristoylamino)-1-phenyl-1-propanol on the regulation of Sertoli cell function. *J. Androl.* **20**(5), 619–625.
203. Grataroli, R., Boussouar, F., and Benahmed, M. (2000). Role of sphingosine in the tumor necrosis factor alpha stimulatory effect on lactate dehydrogenase A expression and activity in porcine Sertoli cells. *Biol. Reprod.* **63**(5), 1473–1481.
204. Ziulkoski, A. L., Zimmer, A. R., Zanettini, J. S., Trugo, L. C., and Guma, F. C. (2001). Synthesis and transport of different sphingomyelin species in rat Sertoli cells. *Mol. Cell Biochem.* **219**(1–2), 57–64.